



Kermanshah  
University Of  
Medical Sciences

In the name of GOD



# Journal Club Presentation

Supervisor: Dr Mohammadzadeh

Student: Ali Vafaei

May 2023



## Article

# Small RNAs are modified with N-glycans and displayed on the surface of living cells

Ryan A. Flynn,<sup>1,10,11,12,\*</sup> Kayvon Pedram,<sup>1</sup> Stacy A. Malaker,<sup>1</sup> Pedro J. Batista,<sup>2</sup> Benjamin A.H. Smith,<sup>3</sup> Alex G. Johnson,<sup>4</sup> Benson M. George,<sup>5</sup> Karim Majzoub,<sup>6,7</sup> Peter W. Villalta,<sup>8</sup> Jan E. Carette,<sup>6</sup> and Carolyn R. Bertozzi<sup>1,9,\*</sup>

<sup>1</sup>Department of Chemistry, Stanford University, Stanford, CA, USA

<sup>2</sup>Laboratory of Cell Biology, Center for Cancer Research, National Cancer Institute, National Institutes of Health, Bethesda, MD, USA

<sup>3</sup>Department of Chemical and Systems Biology and ChEM-H, Stanford University, Stanford, CA, USA

<sup>4</sup>Department of Chemical and Systems Biology, Stanford University, Stanford, CA, USA

<sup>5</sup>Department of Cancer Biology, Stanford University, Stanford, CA, USA

<sup>6</sup>Department of Microbiology and Immunology, Stanford University, Stanford, CA, USA

<sup>7</sup>IGMM, CNRS, University of Montpellier, Montpellier, France

<sup>8</sup>Masonic Cancer Center and Department of Medicinal Chemistry, University of Minnesota, Minneapolis, MN, USA

<sup>9</sup>Howard Hughes Medical Institute, Stanford University, Stanford, CA, USA

<sup>10</sup>Present address: Stem Cell Program, Boston Children's Hospital, Boston, MA, USA

<sup>11</sup>Present address: Department of Stem Cell and Regenerative Biology, Harvard University, Cambridge, MA, USA

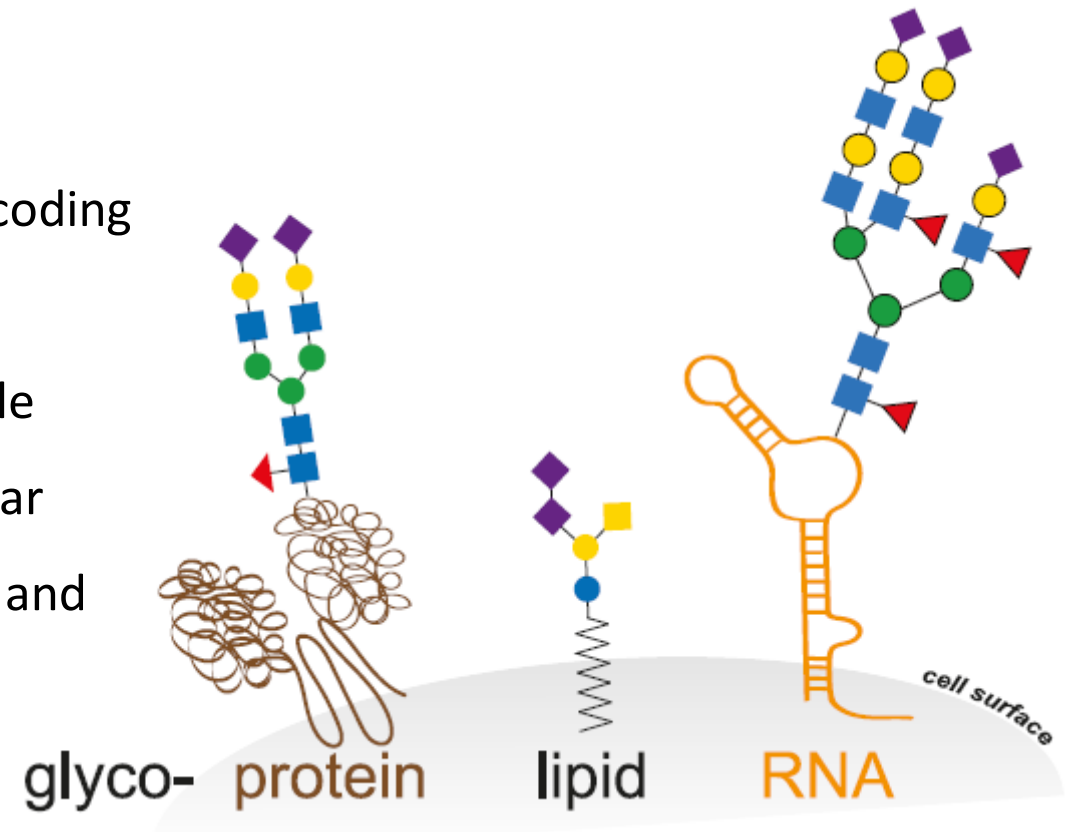
<sup>12</sup>Lead contact

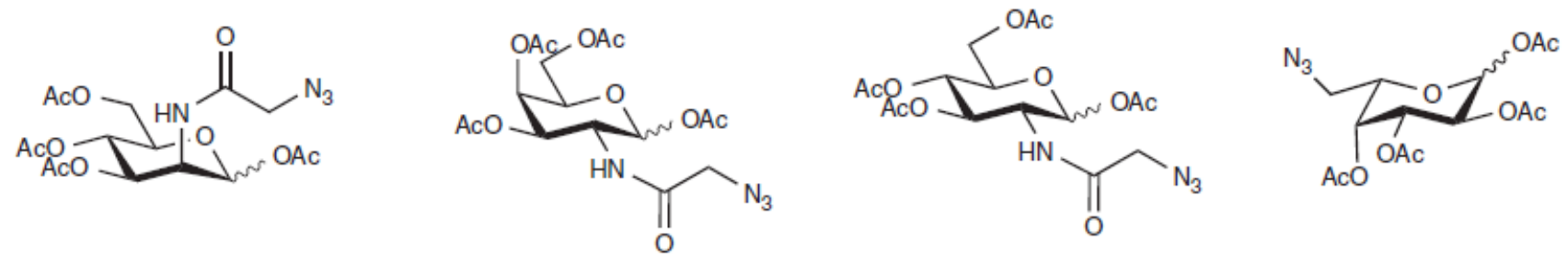
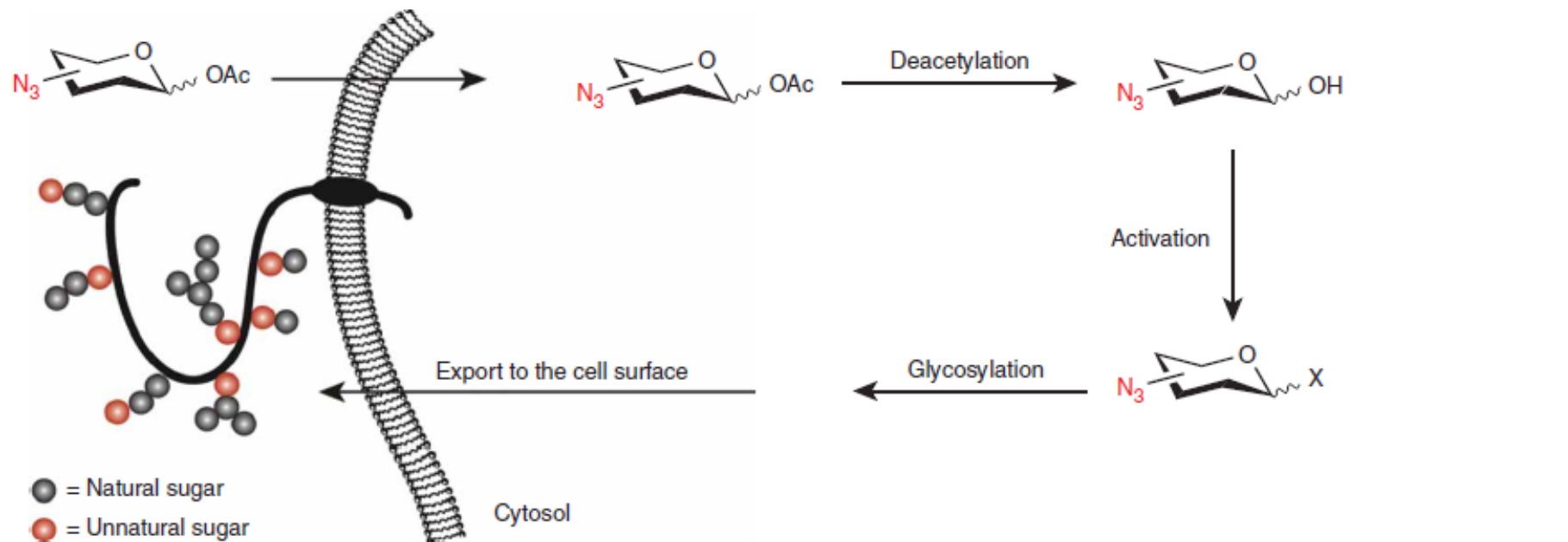
\*Correspondence: [ryan.flynn@childrens.harvard.edu](mailto:ryan.flynn@childrens.harvard.edu) (R.A.F.), [bertozzi@stanford.edu](mailto:bertozzi@stanford.edu) (C.R.B.)

<https://doi.org/10.1016/j.cell.2021.04.023>

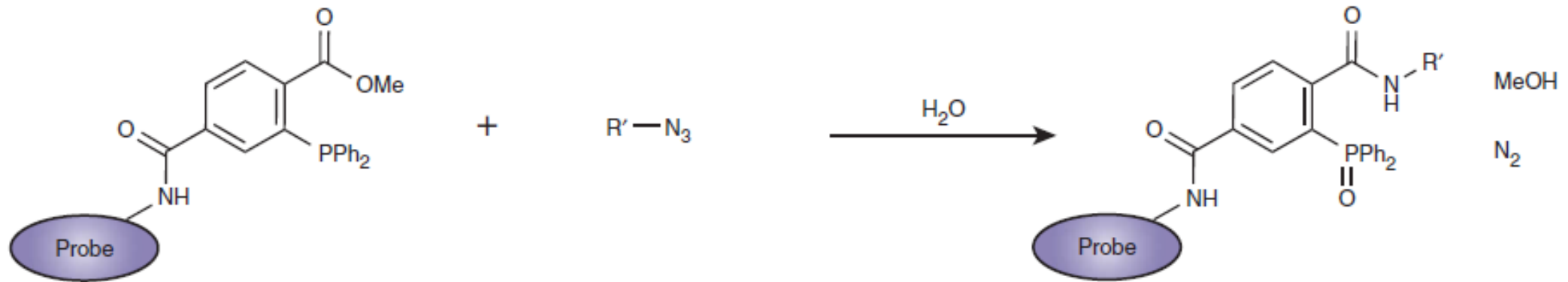
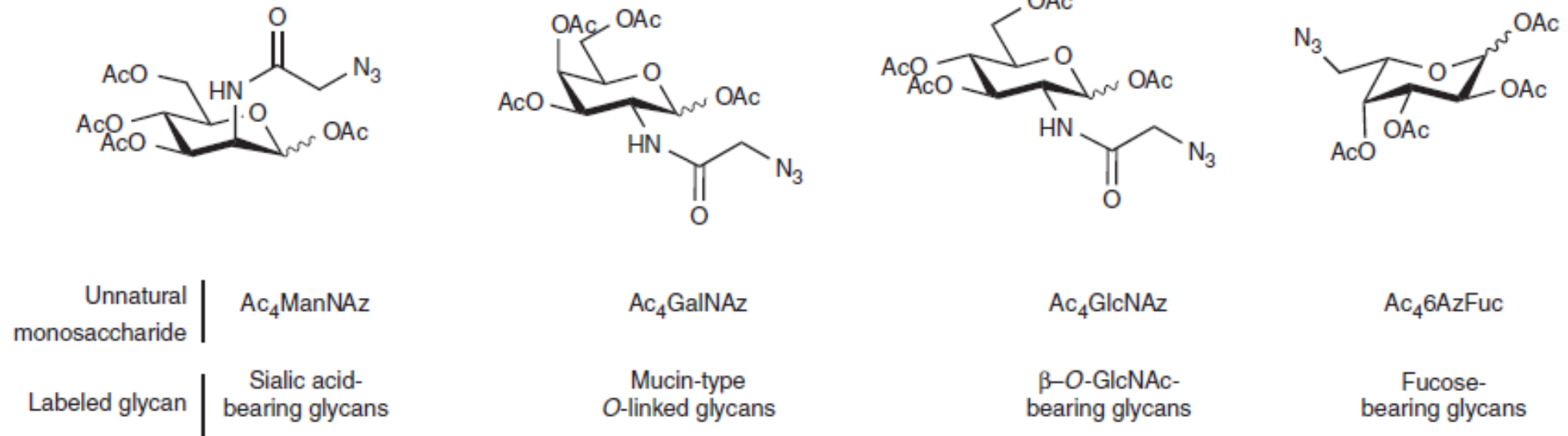
# INTRODUCTION:

- ✓ Glycans are present in every cell studied to date across the kingdoms of life
- ✓ Glycans modify lipids and proteins to mediate inter- and intramolecular interactions across all domains of life.
- ✓ By chemical and biochemical approaches, defined small noncoding RNAs can be as third scaffold for glycosylation.
- ✓ the cellular role for RNA is more complex than that of a simple messenger. For instance, RNAs function as scaffolds, molecular decoys, enzymes, and network regulators across the nucleus and cytosol





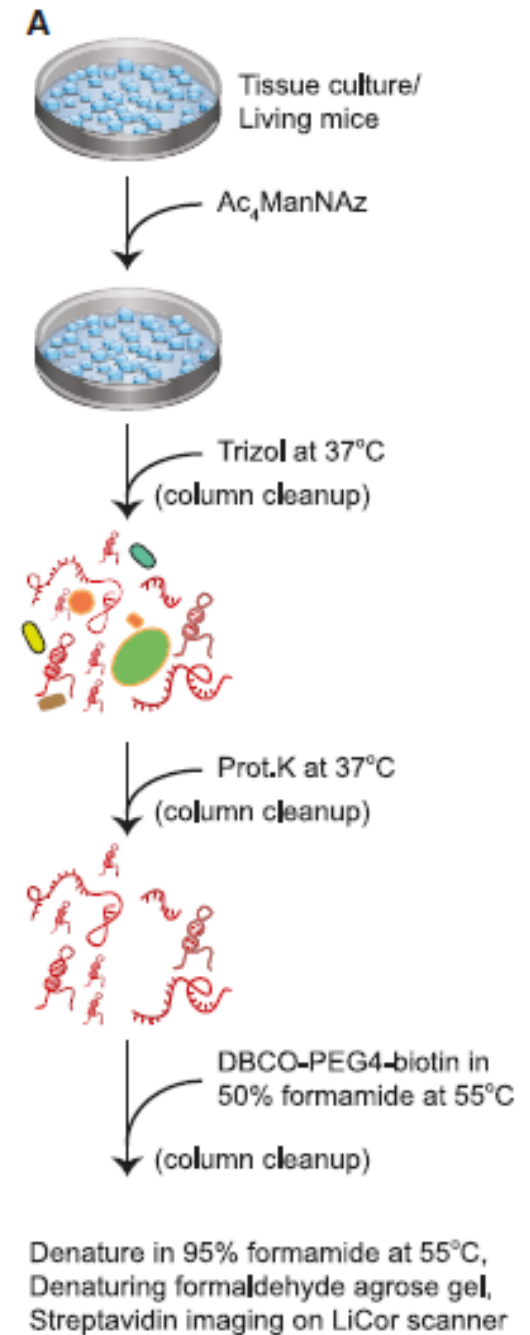
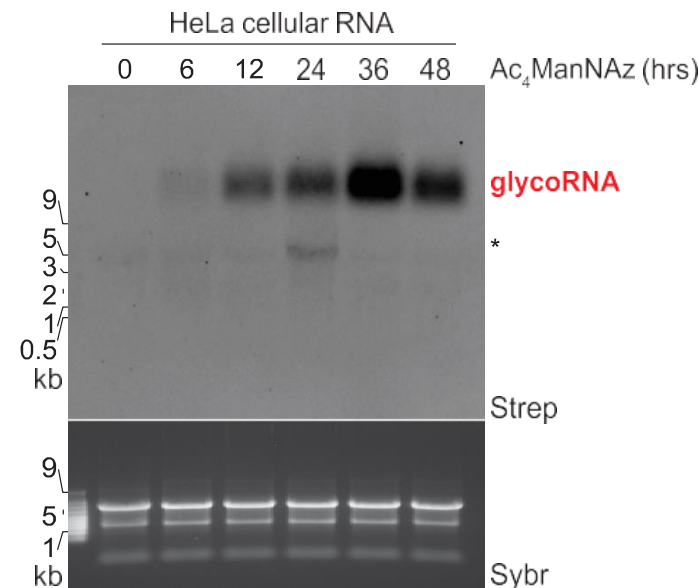
- ✓ In this strategy, metabolically label cells or animals with precursor sugars functionalized with a clickable azide group. Once incorporated into cellular glycans, the azidosugars enable bioorthogonal reaction with a biotin probe for enrichment, identification, and visualization.



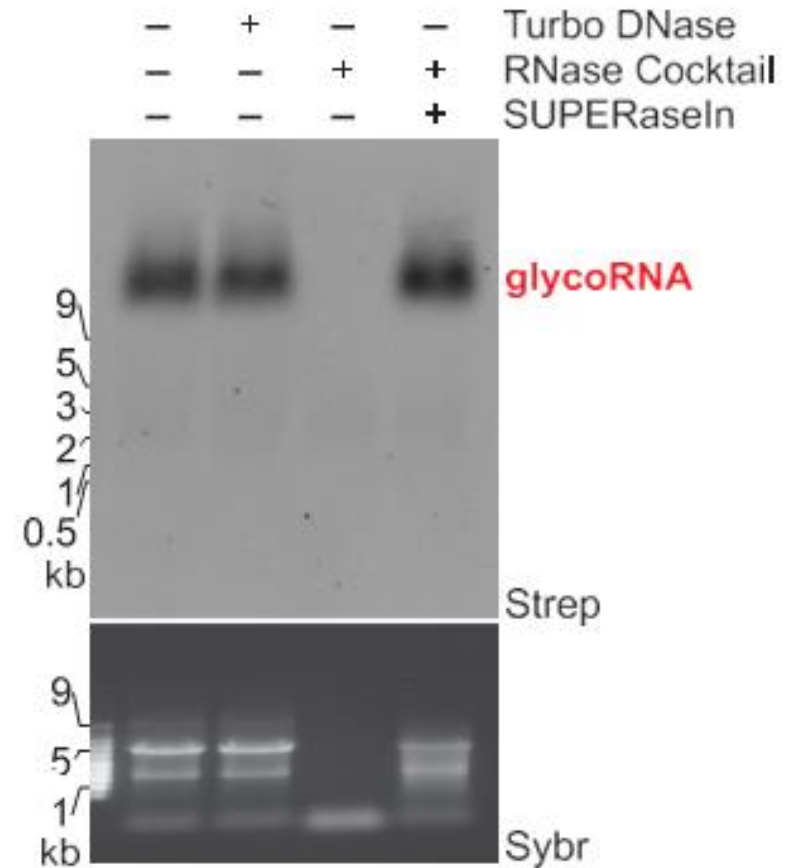
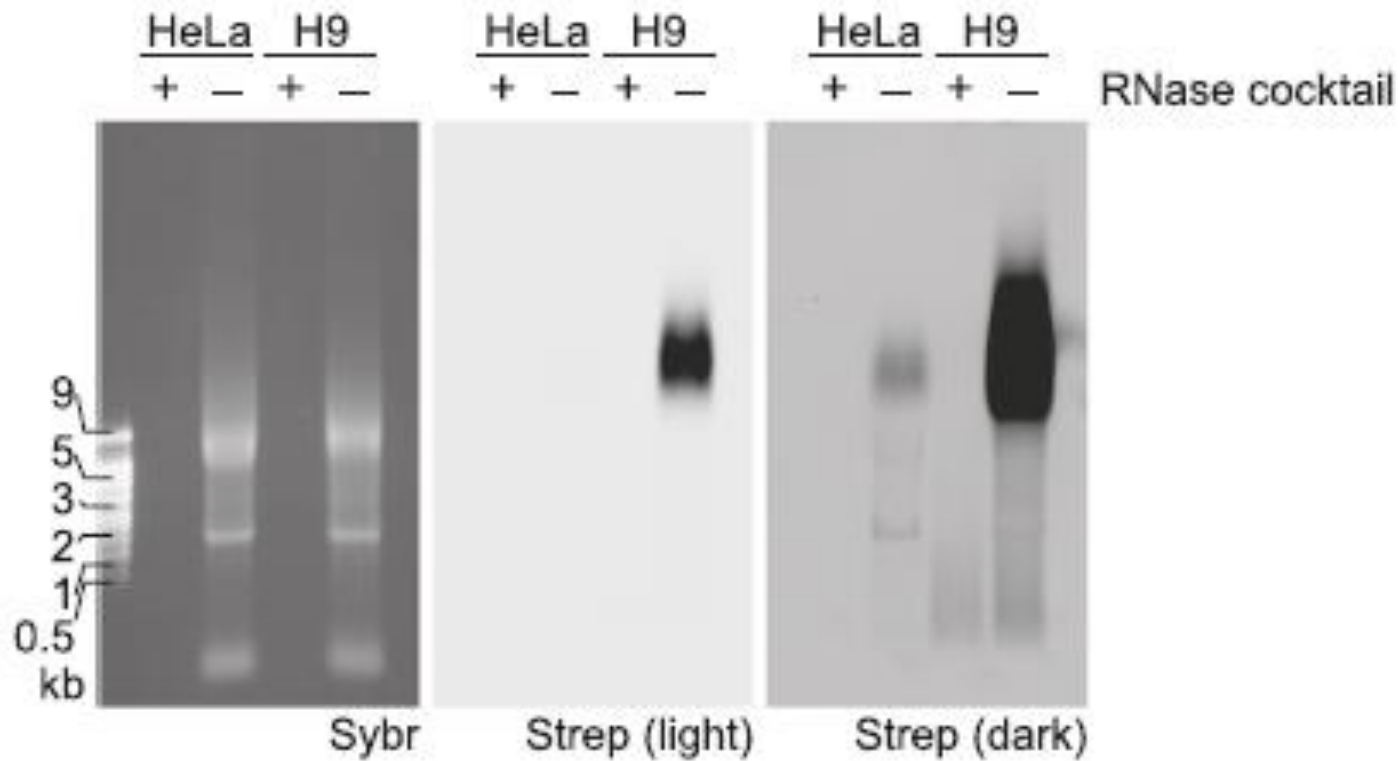
✓ In this strategy, we metabolically label cells or animals with precursor sugars functionalized with a clickable azide group. Once incorporated into cellular glycans, the azidosugars enable bioorthogonal reaction with a biotin probe for enrichment, identification, and visualization.

# A glycan metabolic reporter is incorporated into cellular RNA

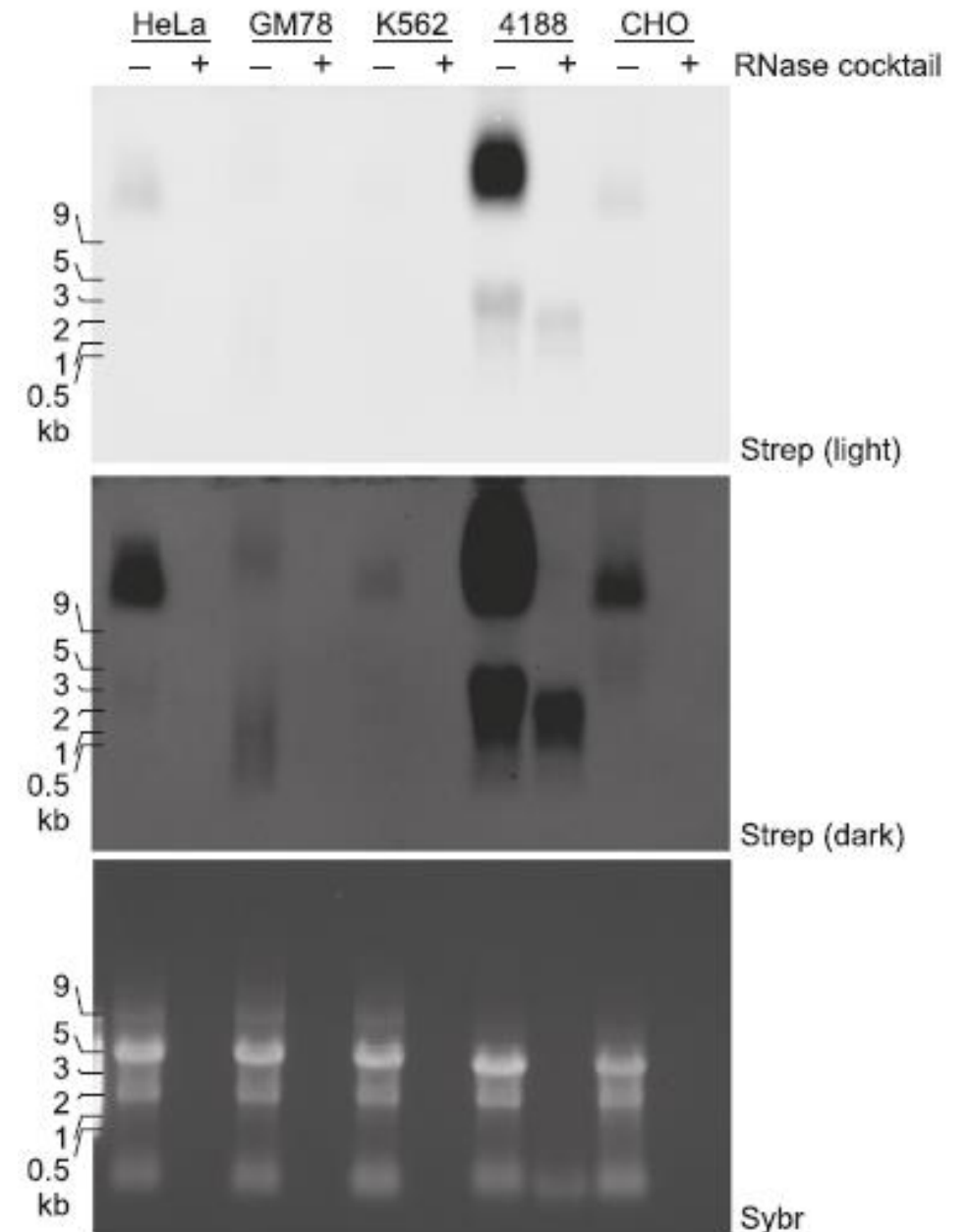
- ✓ First HeLa cell treated by 100 mM Ac<sub>4</sub>ManNAz for up to 48 h and then RNA is extracted with warm TRIzol method.
- ✓ To visualize azide-labeled components, RNA samples added to dibenzocyclooctyne-biotin (DBCO-biotin) in denaturing conditions (50% formamide) at 55°C, subsequently separated by denaturing gel electrophoresis and analyzed by blotting



- ✓ treatment of RNA from Ac4ManNAz-labeled HeLa cells with DNase did not affect the glycoRNA signal, whereas treatment with an RNase cocktail efficiently digested the total RNA as well as the biotinylated glycoRNA

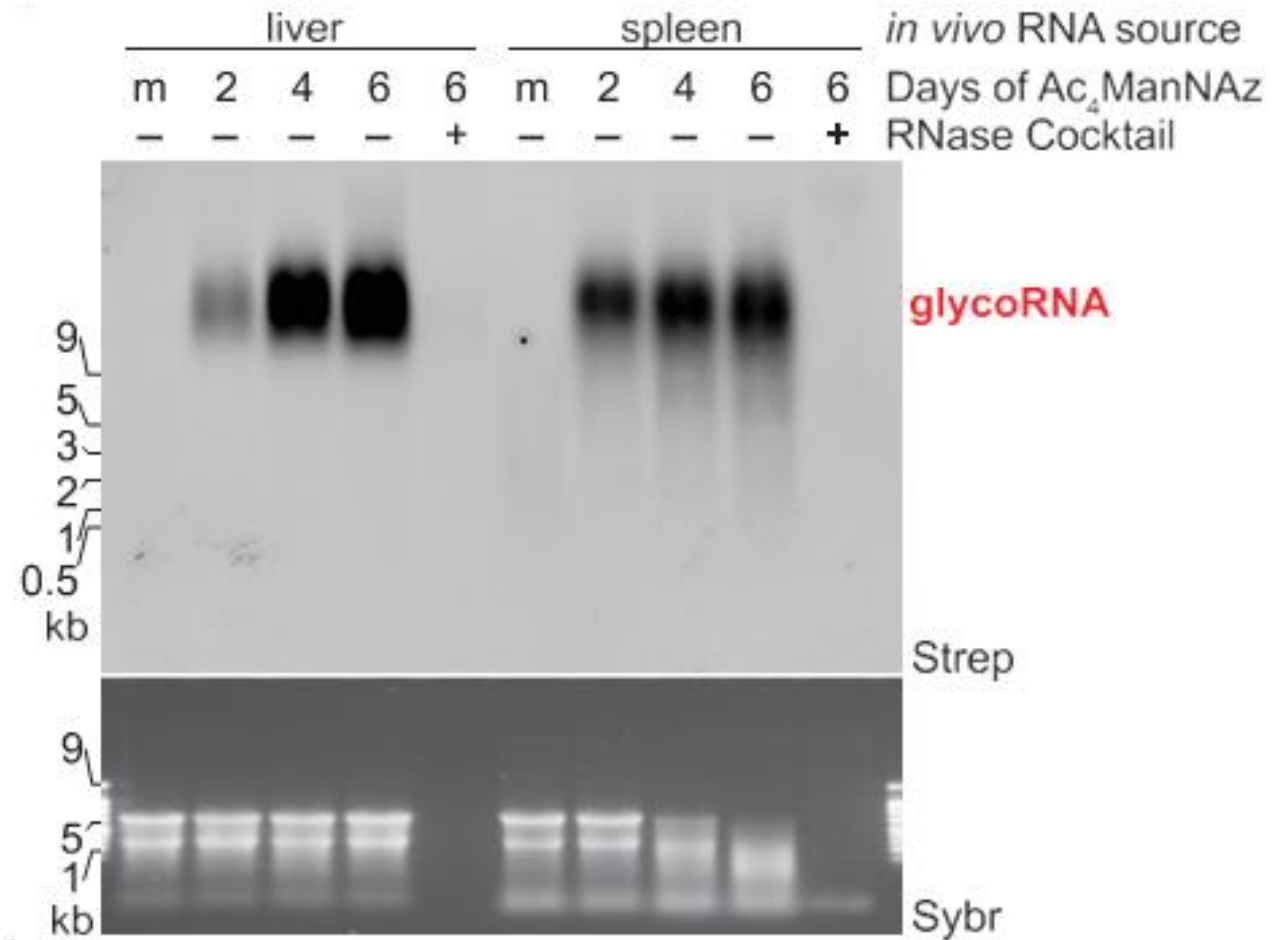


- ✓ Human embryonic stem cells (H9), a human myelogenous leukemia line (K562), a human lymphoblastoid cell line (GM12878), a mouse T cell acute lymphoblastic leukemia cell line (T-ALL 4188), and Chinese hamster ovary cells (CHO) all showed evidence of the presence of glycoRNA.
- ✓ H9 and 4188 cells showed significantly more labeling with Ac4ManNAz per mass of total RNA than other cell types



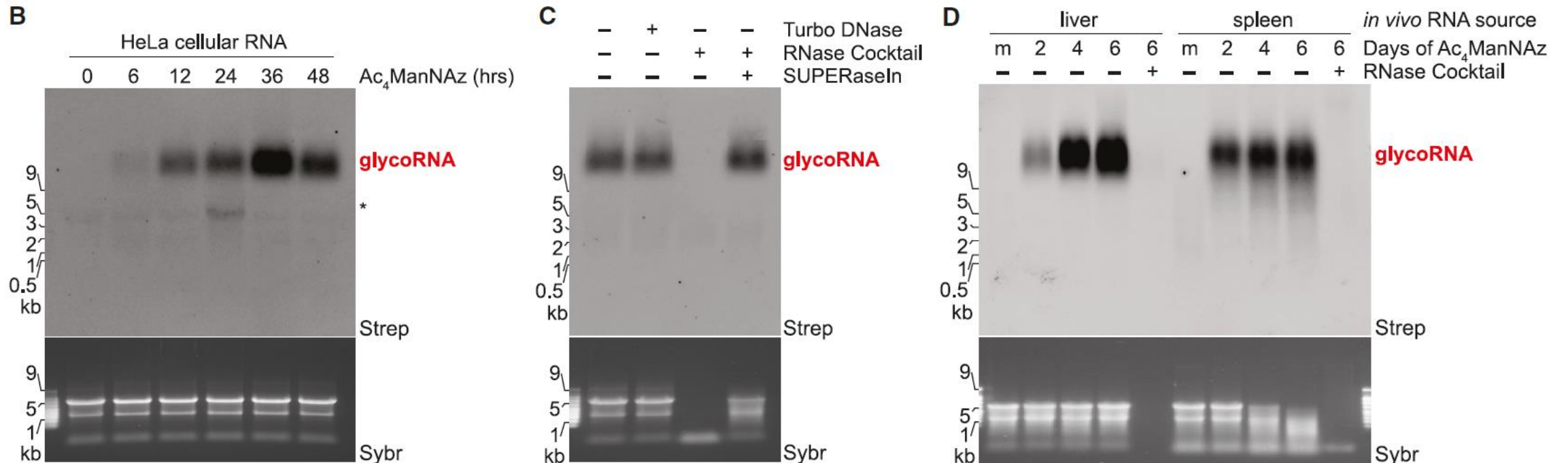


- ✓ Next, assessed this labeling in vivo.
- ✓ To this end, performed intraperitoneal injections of Ac4ManNAz into mice for 2, 4, or 6 days.
- ✓ In the liver and spleen, the organs that yielded enough total RNA for analysis, we observed dose-dependent and RNase-sensitive Ac4ManNAz labeling of RNAs in the same MW region as glycoRNAs from cultured cells
- ✓ These data suggest that glycoRNA is not an artifact of tissue culture and occurs broadly across multiple cell and tissue types and at various abundances.

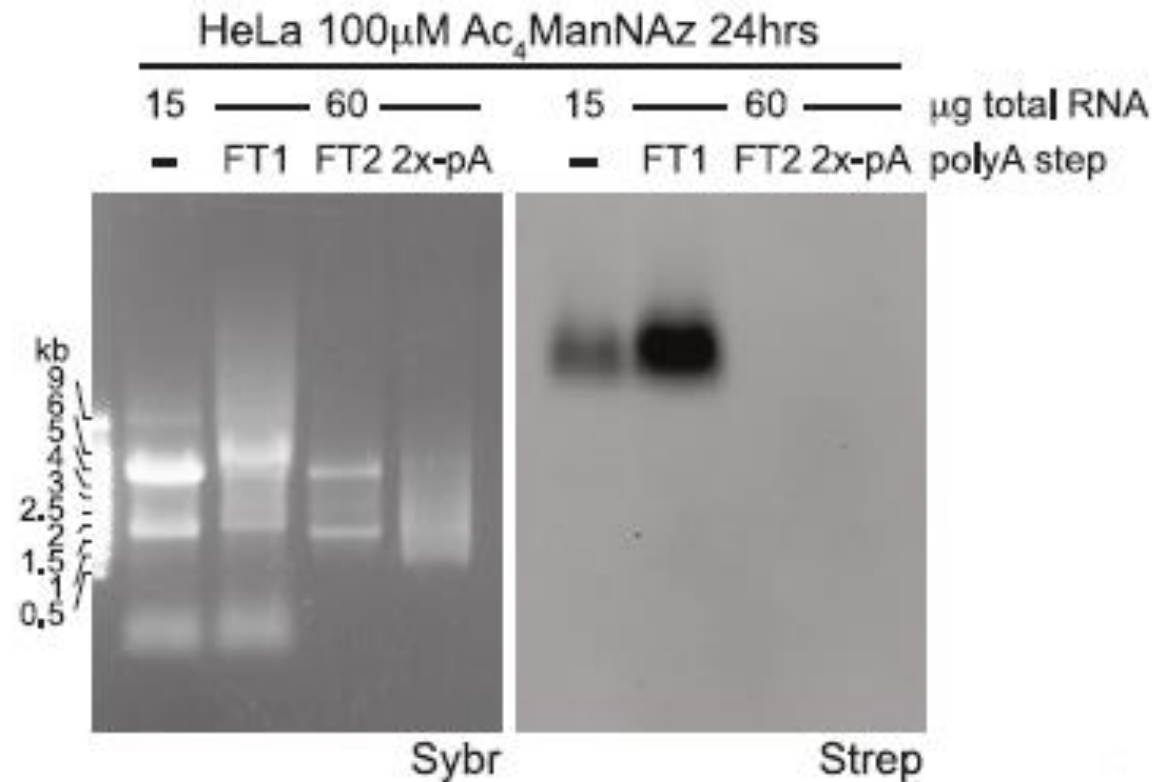
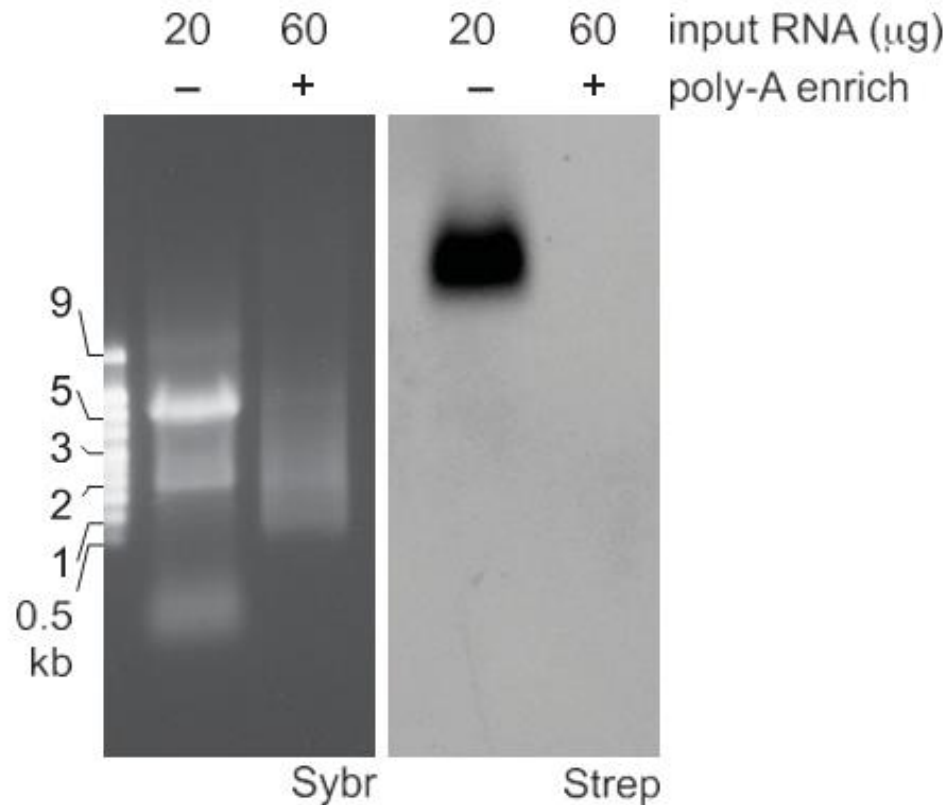


# glycoRNAs are small noncoding RNAs

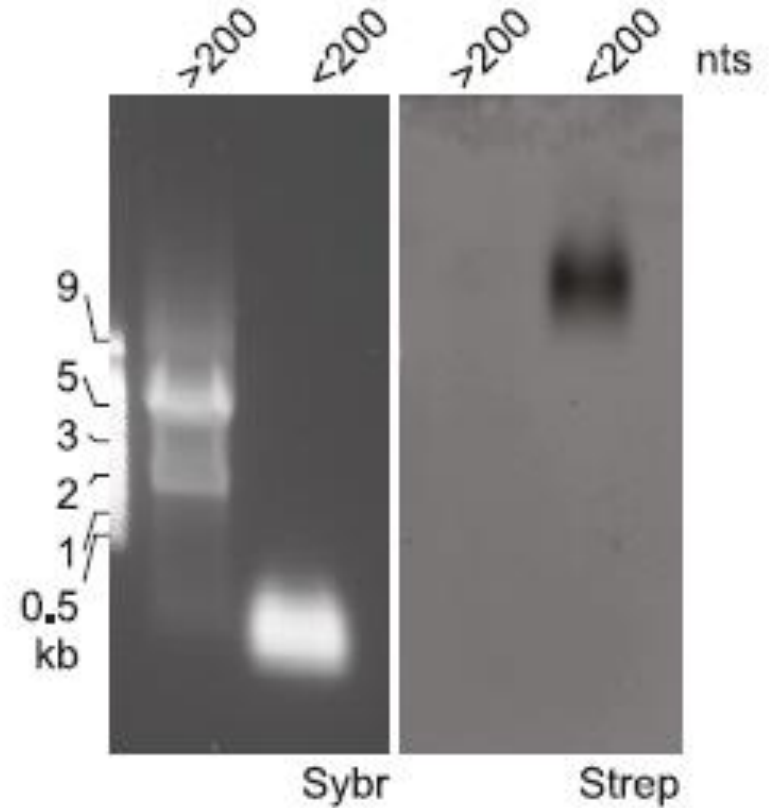
- ✓ Across all cell types and organs tested, glycoRNA was found to migrate very slowly by denaturing agarose gel electrophoresis.



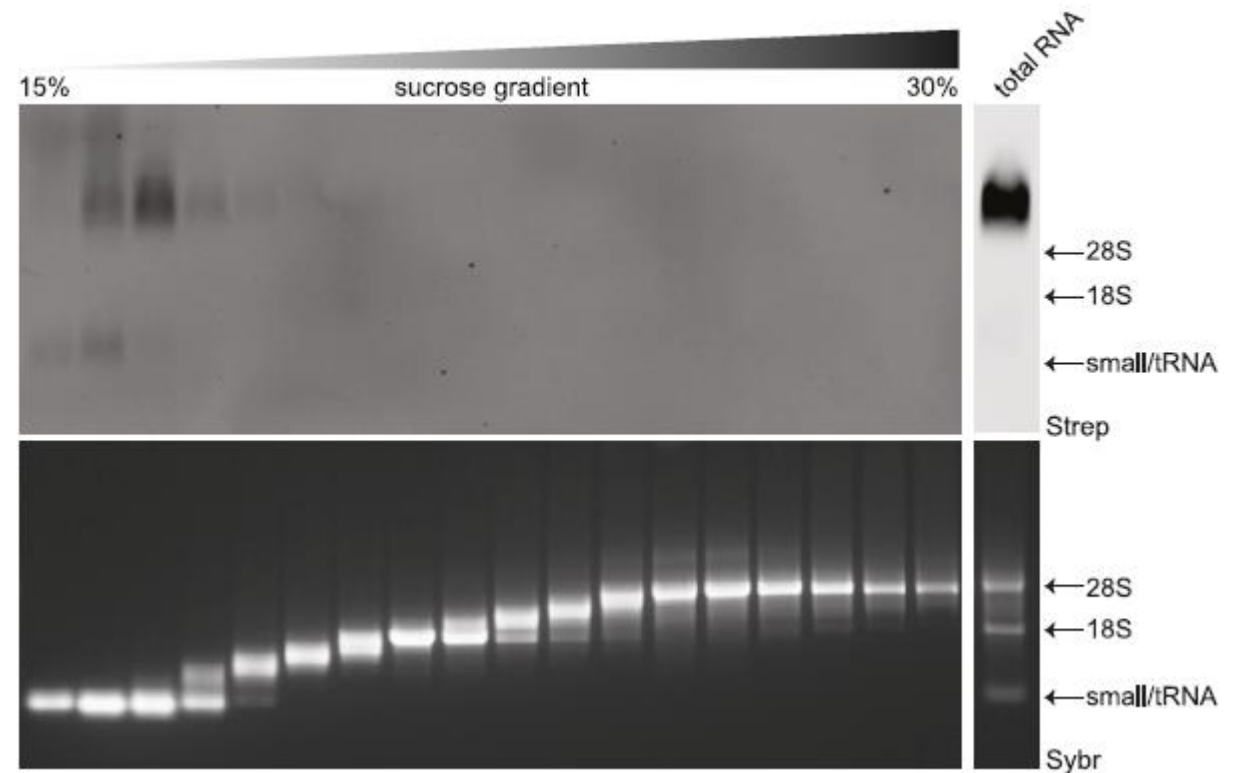
- ✓ We hypothesized that if glycoRNA's are indeed large RNAs, they would likely be polyadenylated (poly-A). However, we were consistently unable to purify glycoRNA from extracted RNA via poly-A enrichment.
- ✓ This was not due to cleavage or degradation of the glycoRNA during the poly-A enrichment procedure.



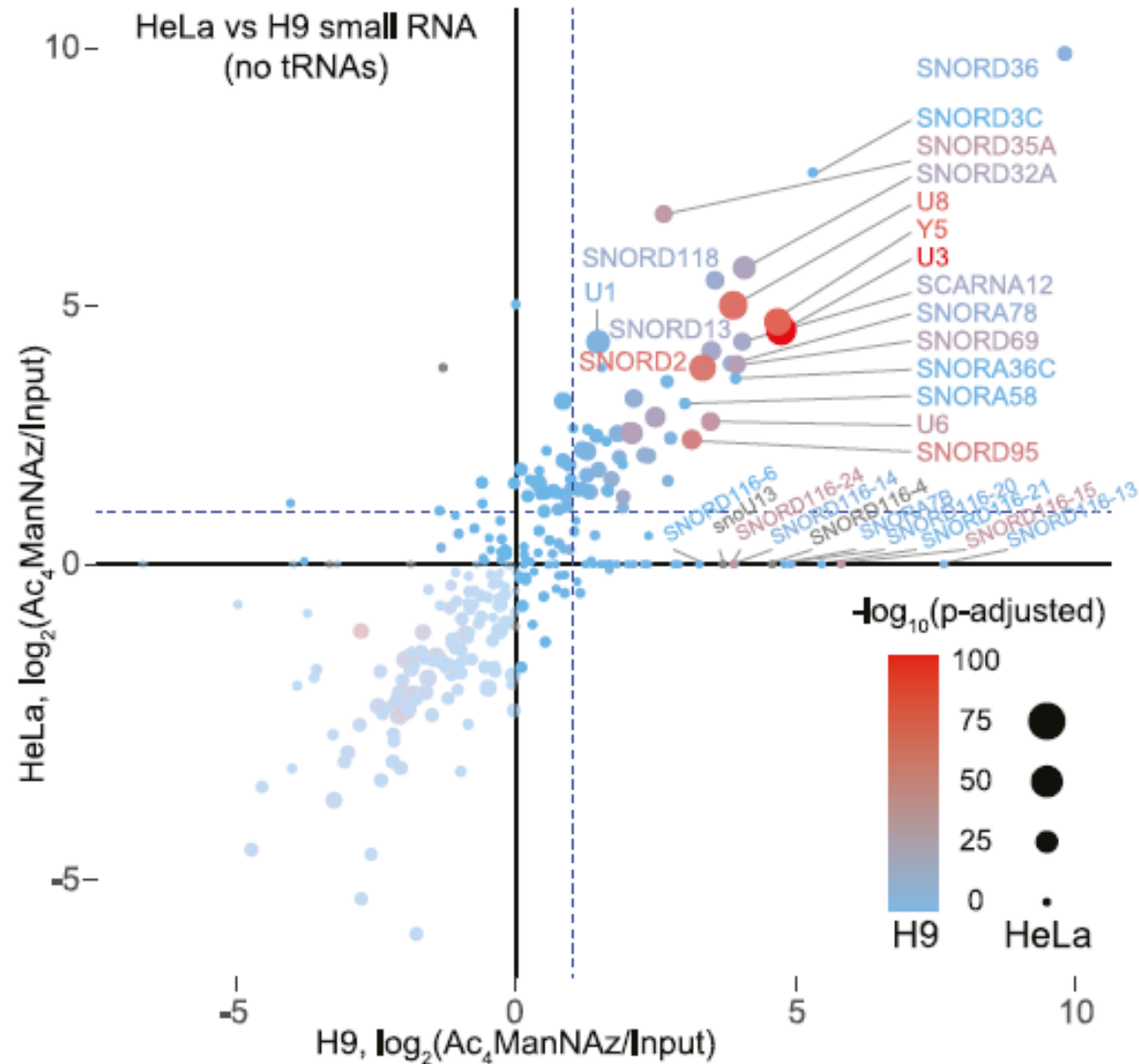
- ✓ As an alternative enrichment strategy, we used a commercial fractionation method that leverages length-dependent RNA precipitation and binding to silica columns to separate out “large” (>200 nt) from “small” (<200 nt) transcripts (STAR Methods). To our surprise, the glycoRNA fractionated exclusively with the small RNA population of total RNA



- ✓ The sucrose gradient robustly separated the major visible RNAs such as small RNAs/tRNA, 18S rRNA, and 28S rRNA
- ✓ glycoRNA's anomalous migratory behavior is caused by its associated glycans



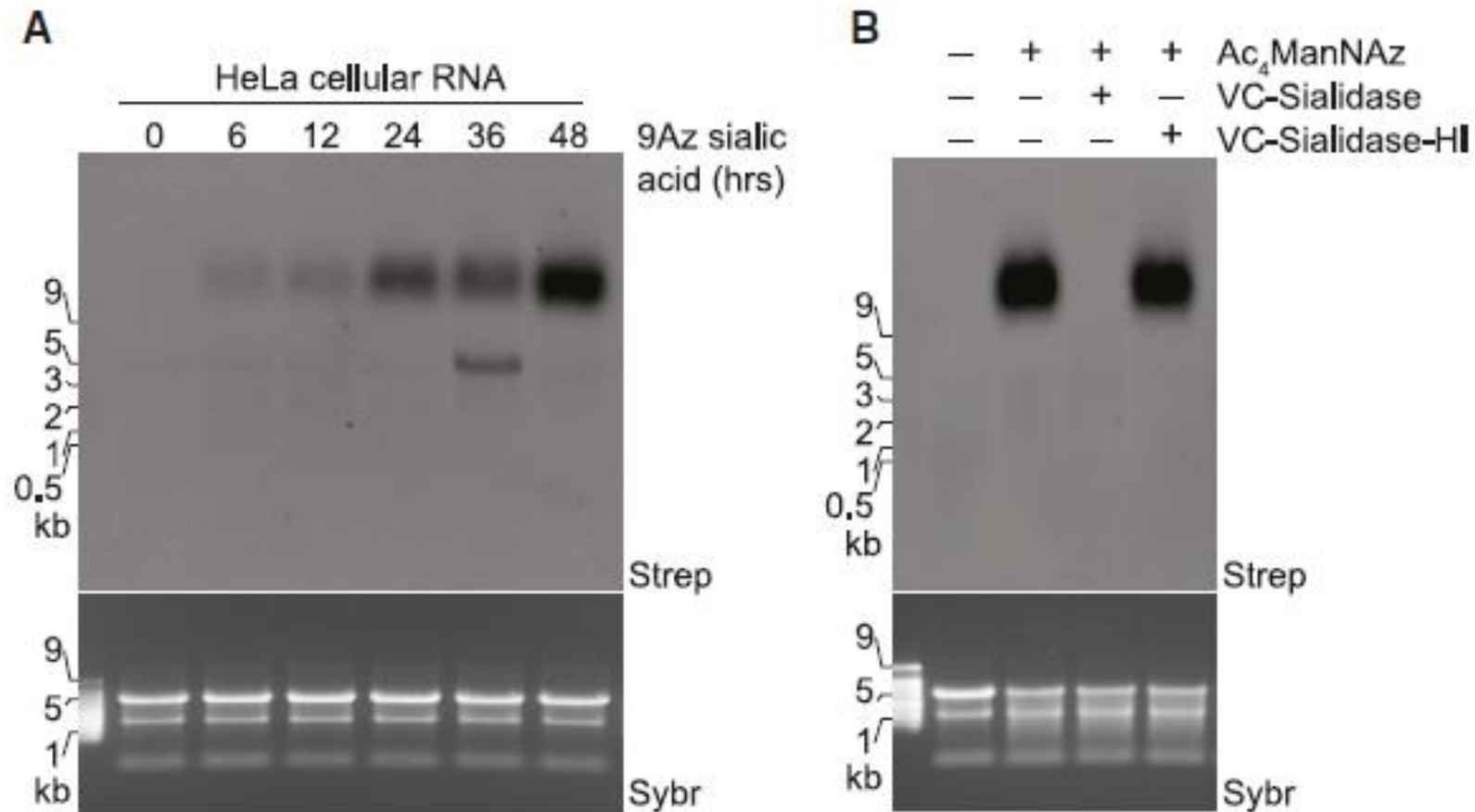
# A common set of transcripts are glycosylated across diverse cell types



## **Label and label-free detection of sialic acid in glycoRNA**

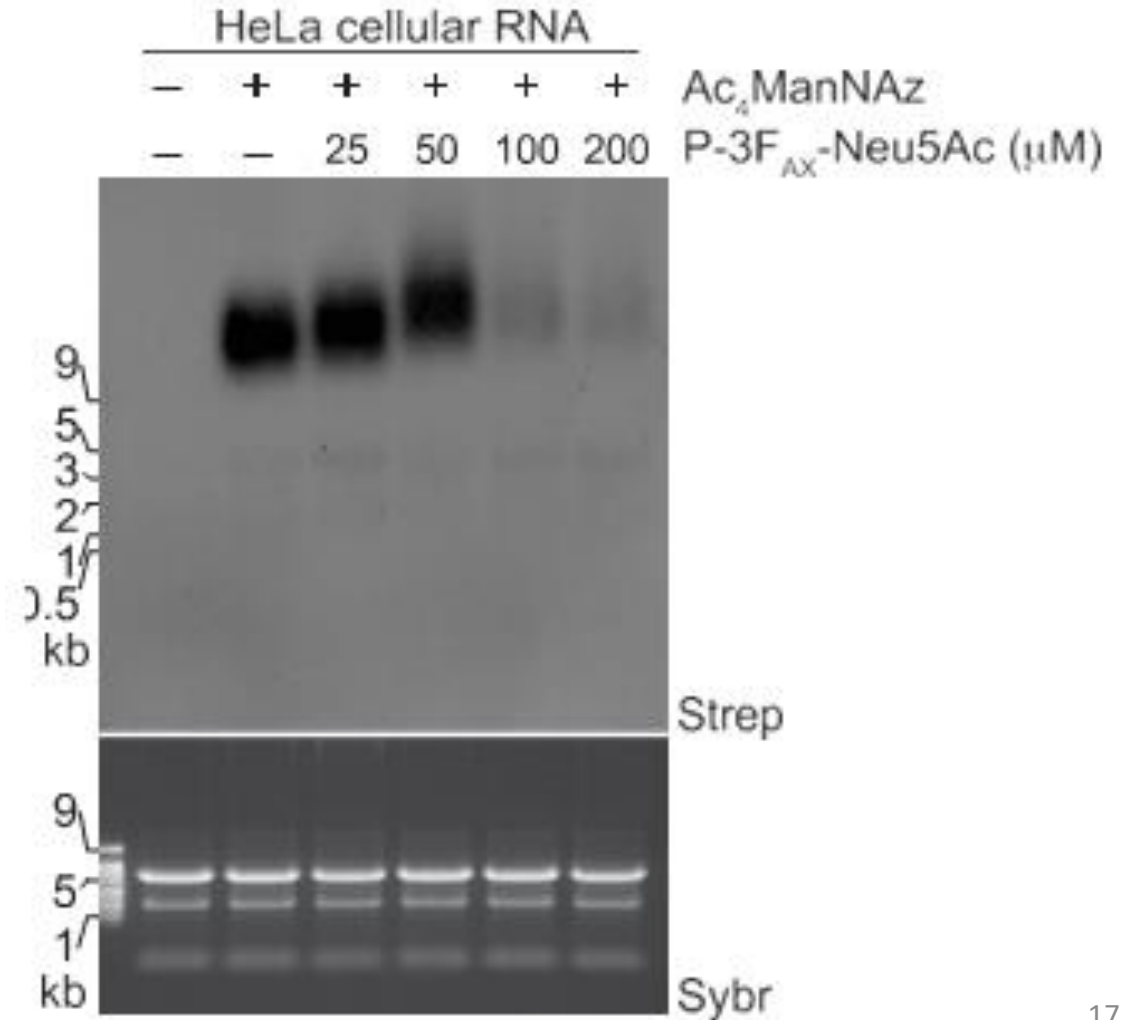
- ✓ **Next step is determination the glycan structures on glycoRNAs**
- ✓ **Use 2 different method: metabolic and non-metabolic**

- ✓ To exclude the possibility that Ac<sub>4</sub>ManNAz is shunted into unexpected metabolic pathways, used 9-azido sialic acid (9Azsialic acid), which is directly converted into CMP-sialic as a metabolic label.

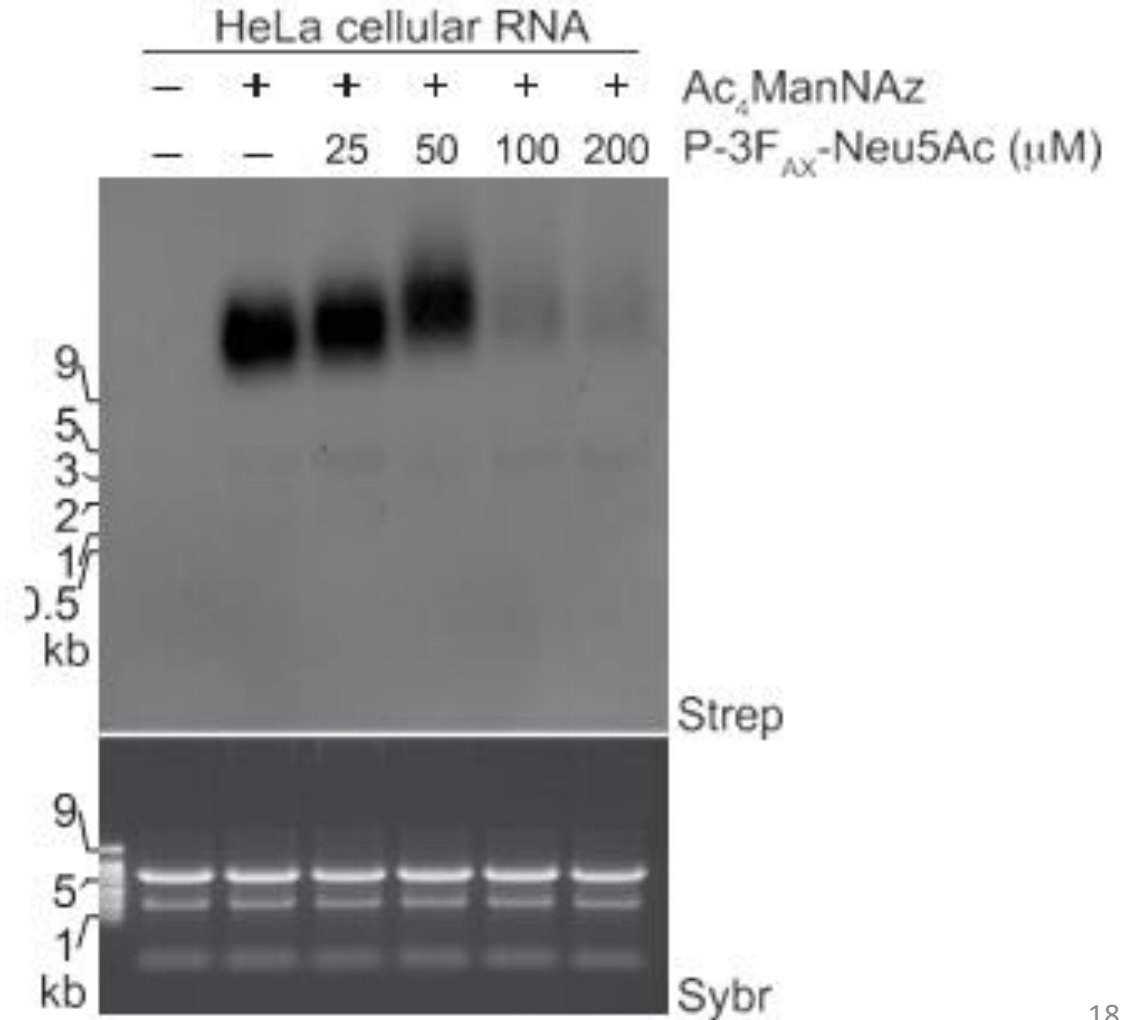




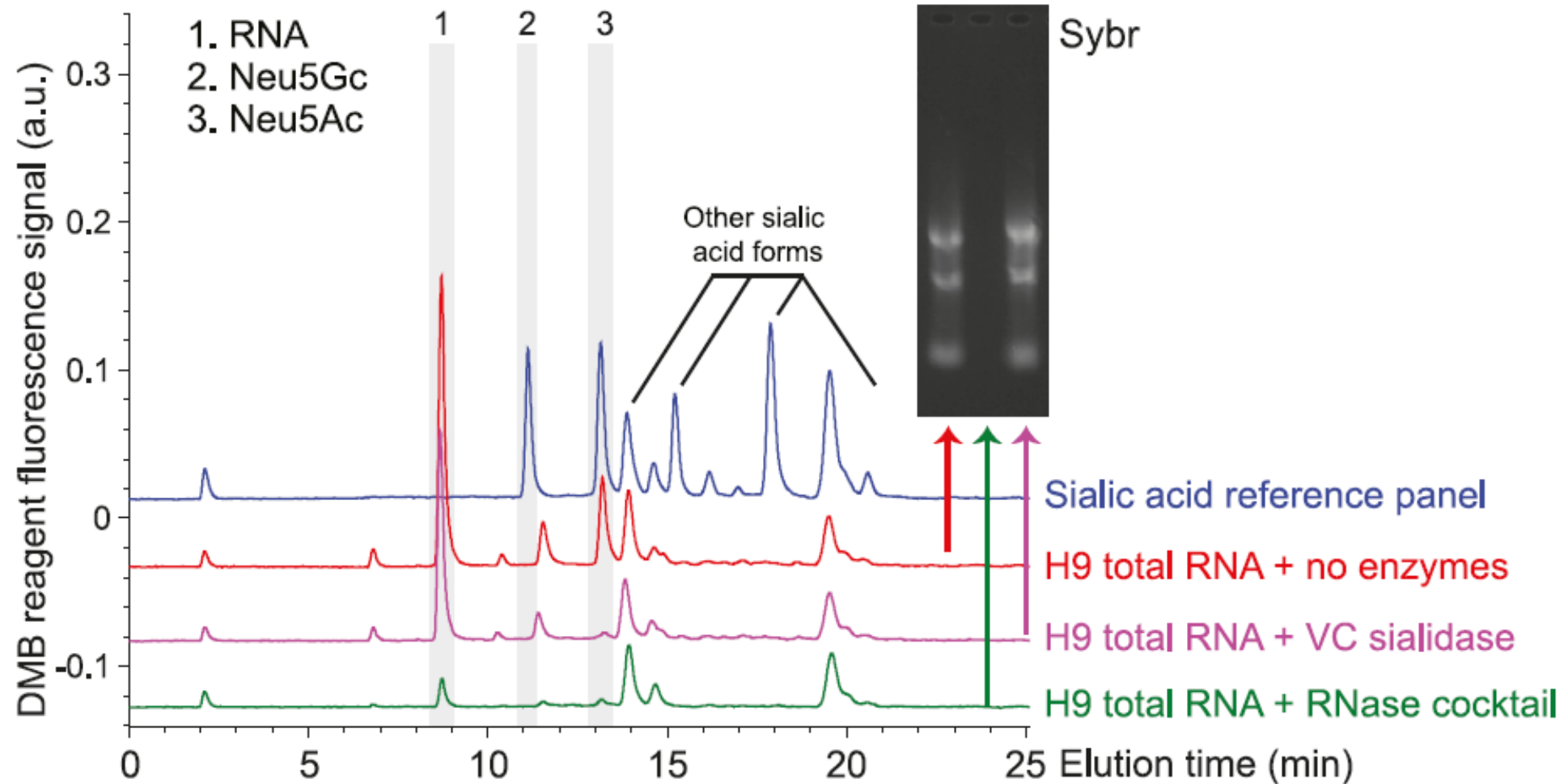
- ✓ assessed the contribution of canonical sialic acid biosynthesis enzymes through the use of P-3F<sub>AX</sub>-Neu5Ac, a cell-permeable metabolic inhibitor of sialoside biosynthesis



- ✓ assessed the contribution of canonical sialic acid biosynthesis enzymes through the use of P-3F<sub>AX</sub>-Neu5Ac, a cell-permeable metabolic inhibitor of sialoside biosynthesis

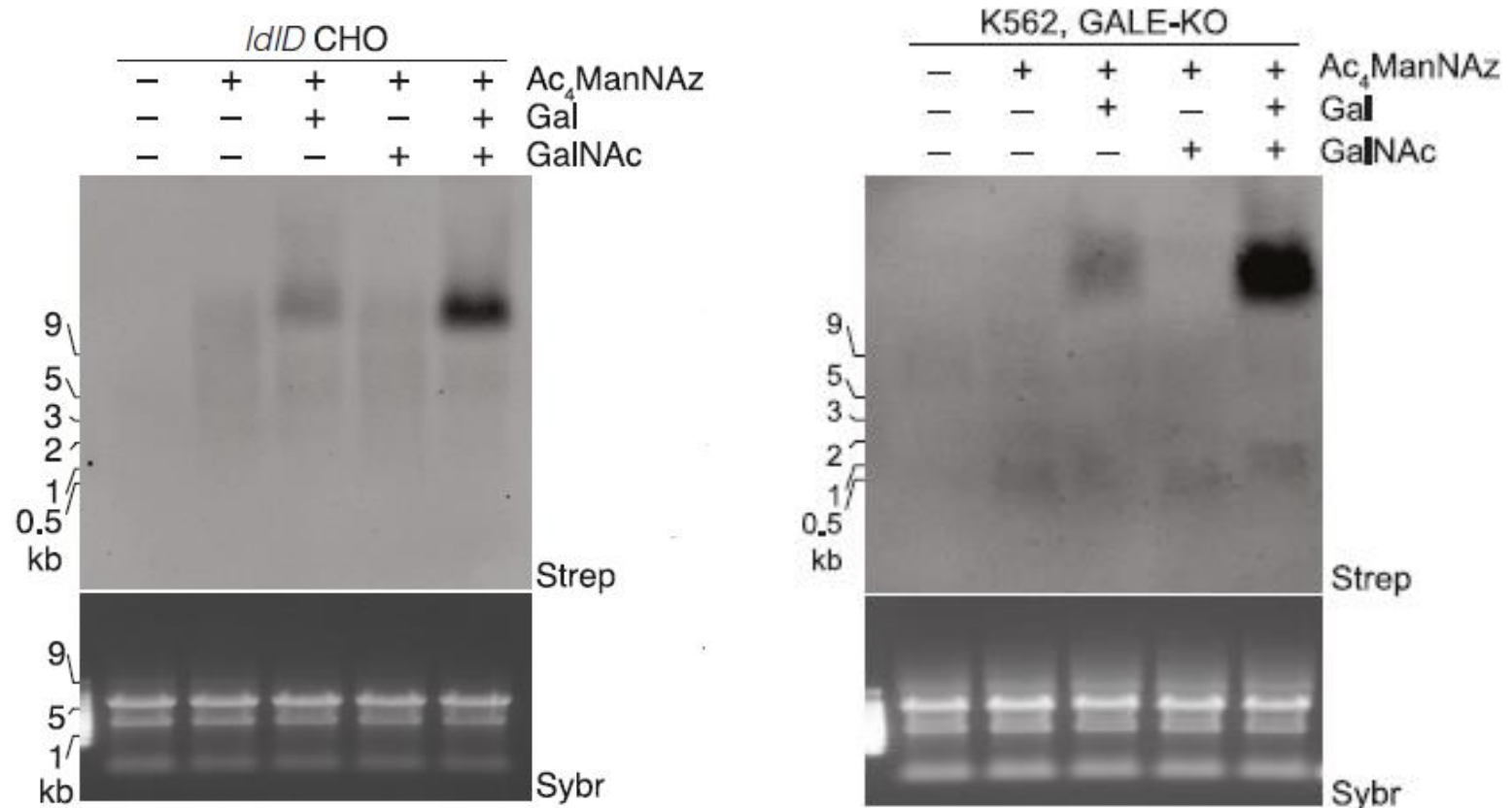


- ✓ To confirm that glycoRNAs are sialylated, used an independent method not relying on metabolic reporters
- ✓ The fluorogenic 1,2-diamino-4,5-methylenedioxybenzene (DMB) probe is used to derivatize free sialic acids for detection and quantitation by high-performance liquid chromatography (HPLC)-fluorescence

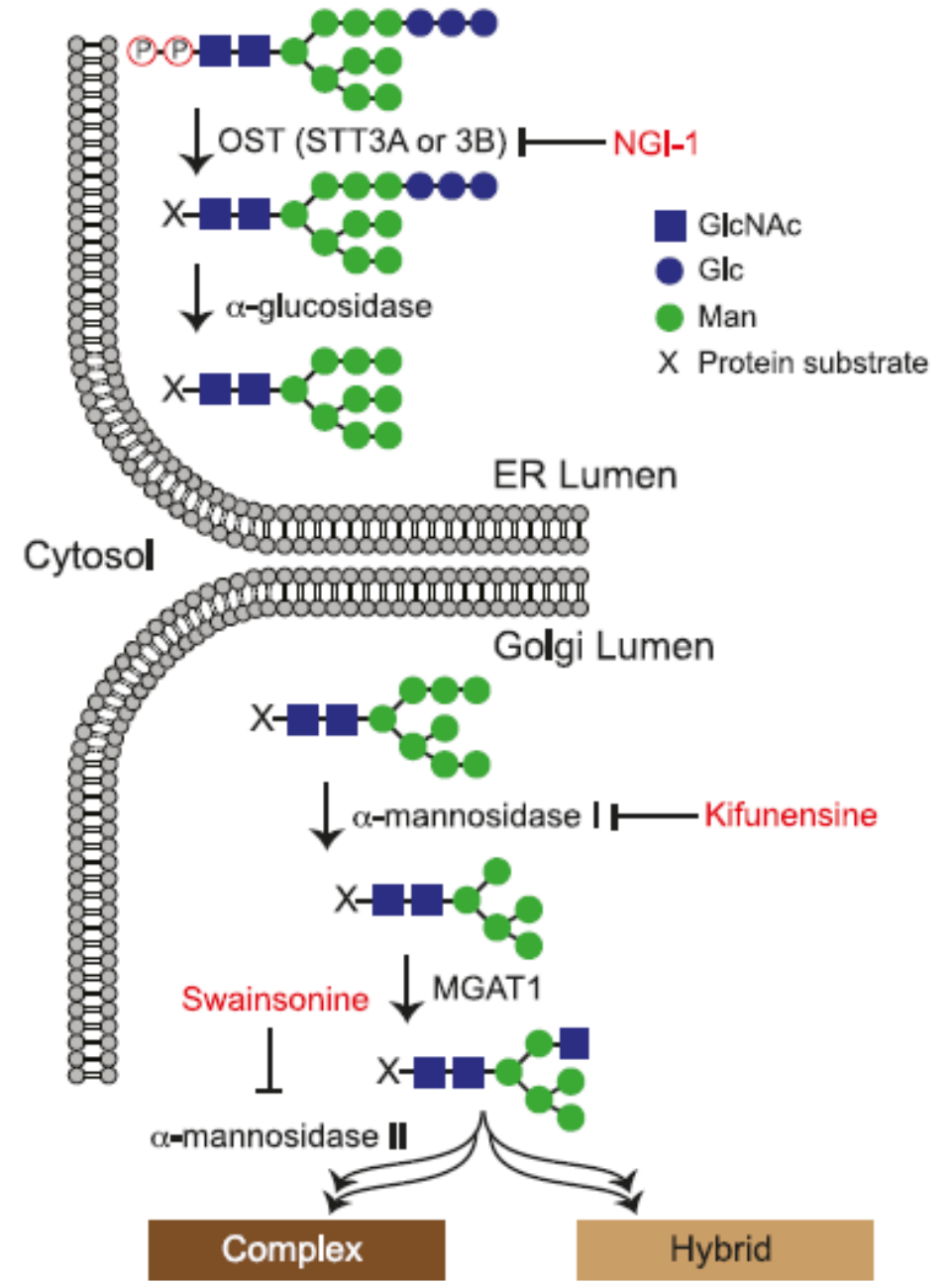
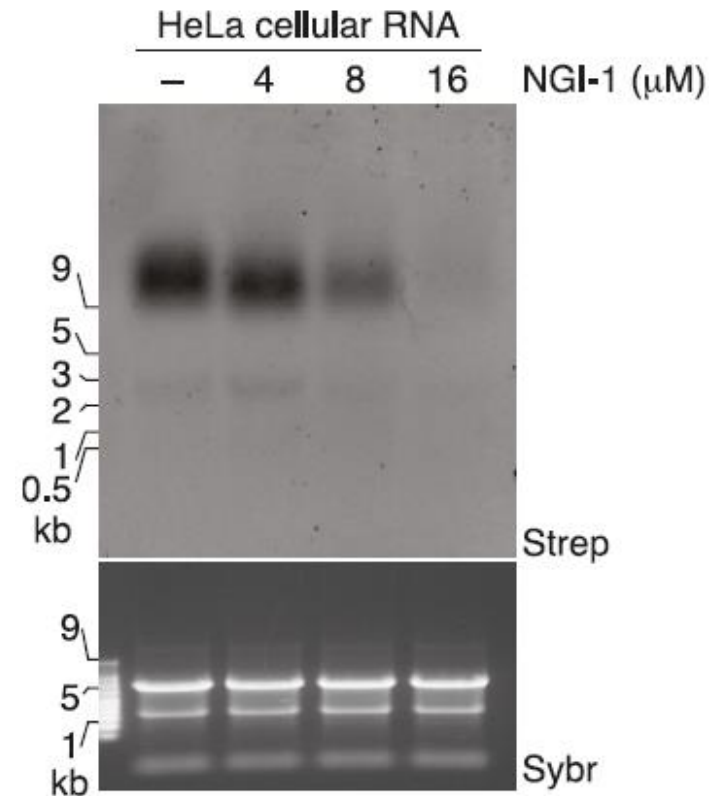


## Canonical N-glycan biosynthetic machinery contributes to glycoRNA production

- ✓ There are two main classes of glycans on proteins, N- and O-glycans, and both can be sialylated
- ✓ The Id1D mutant CHO cell line lacks the ability to interconvert GlcNAc into GalNAc
- ✓ human K562 cell line with a CRISPR-Cas9 targeted KO of UDP-galactose-4-epimerase (GALE), which mimics the phenotype of the Id1D CHO cell line

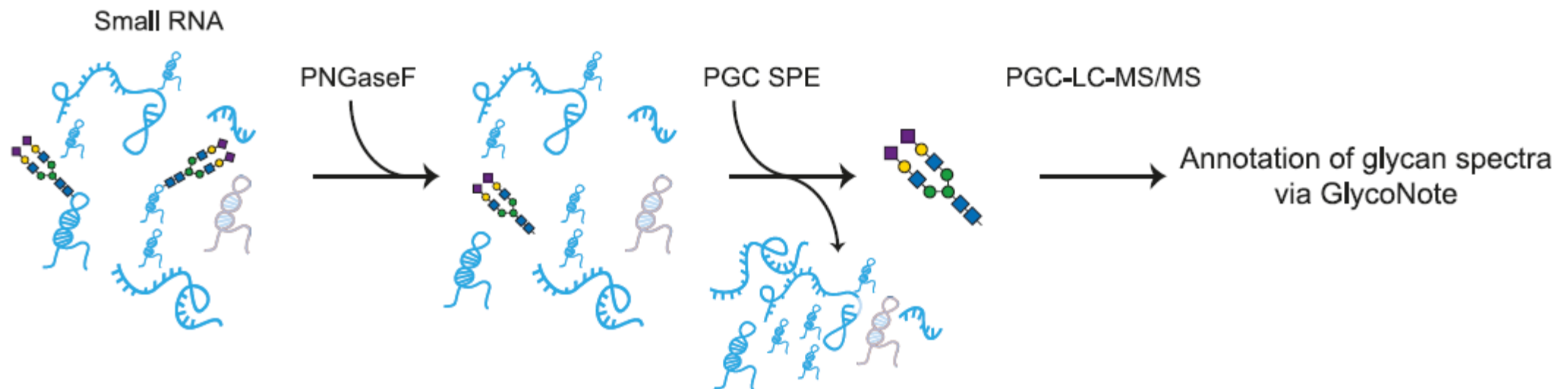


- ✓ Oligosaccharyltransferase (OST) mediates protein N-glycosylation by transferring a 14-sugar glycan to asparagine residues on nascent polypeptides during their translocation through the Sec/translocon
- ✓ tested the effect of NGI-1, a specific and potent small molecule inhibitor of OST, on glycoRNA production



# Mass spectrometry defines distinct compositions of glycans on RNA

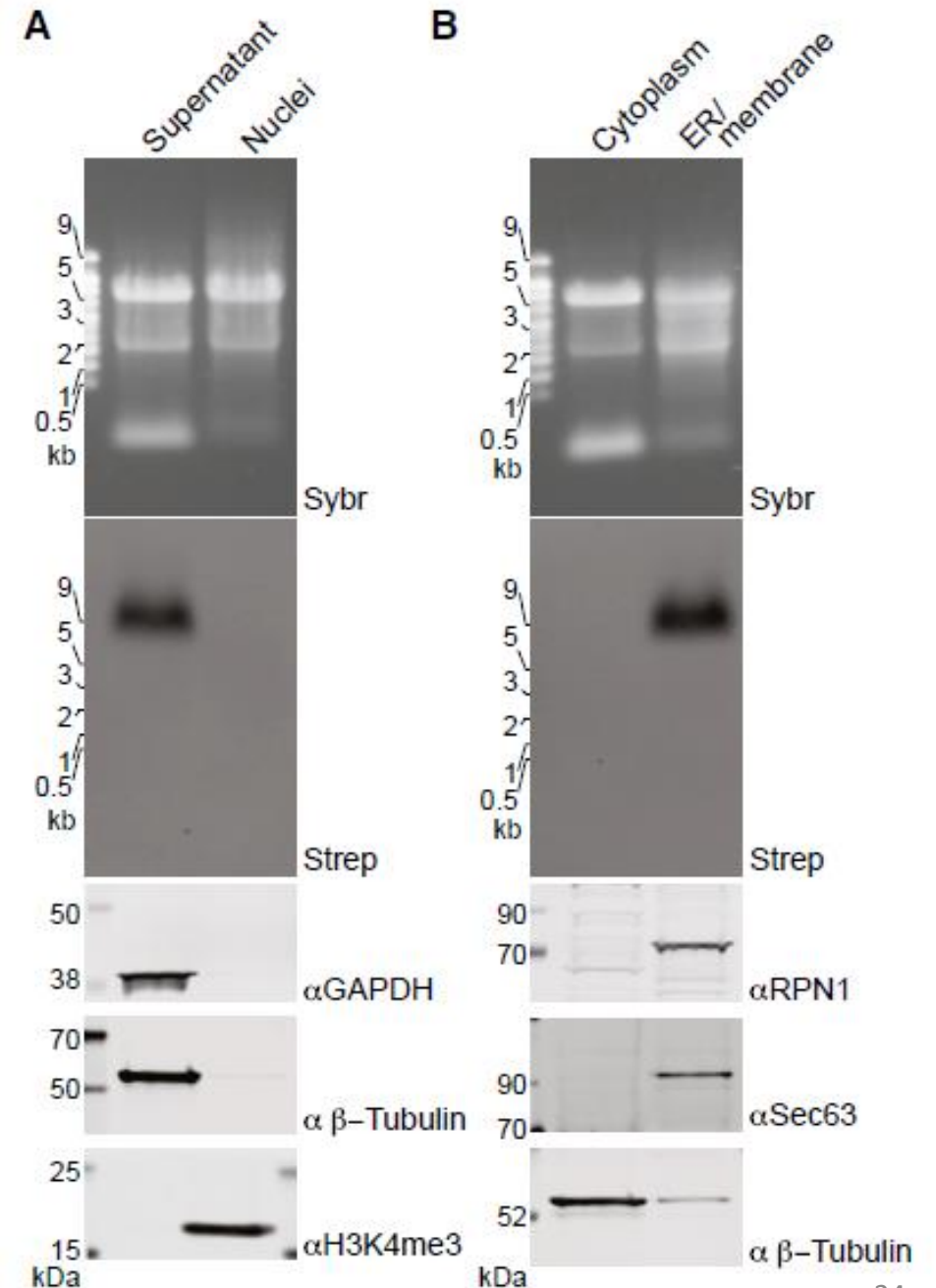
- ✓ To develop a more precise view of the glycoforms associated with RNA, optimized a workflow based on PNGaseF-mediated release of glycans from pools of small RNAs, followed by analysis of those glycans by a porous graphitized carbon-based liquid chromatography MS strategy
- ✓ because the MS- based approach does not require sialic acid for enrichment or visualization, we were able to reveal an expanded set of glycan compositions that are often fucosylated and sometimes asialylated



# glycoRNAs are associated with cellular membrane

- ✓ The localization of Y RNAs has been reported to be mainly cytoplasmic with a minor fraction in the nucleus
- ✓ Other major classes of glycoRNA transcripts such as tRNAs and sn/snoRNAs are classically localized to the soluble cytosol and nucleus, respectively.
- ✓ To determine where glycoRNAs are distributed inside cells, used two biochemical strategies:
  1. Isolates nuclei away from membranous organelles and the cytosol
  2. Separates the soluble cytosolic compartment away from membranous organelles

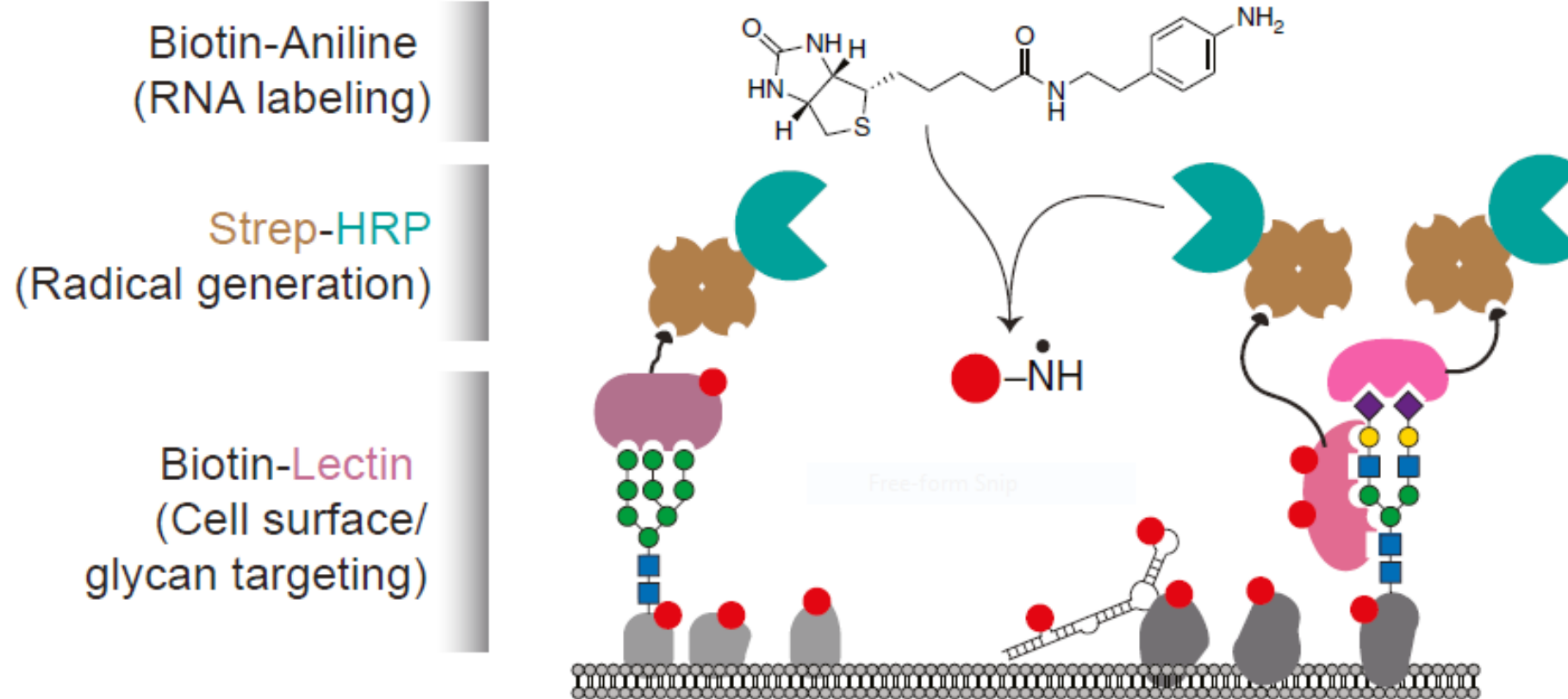
- ✓ The localization of Y RNAs has been reported to be mainly cytoplasmic with a minor fraction in the nucleus
- ✓ Other major classes of glycoRNA transcripts such as tRNAs and sn/snoRNAs are classically localized to the soluble cytosol and nucleus, respectively.
- ✓ To determine where glycoRNAs are distributed inside cells, used two biochemical strategies:
  1. Isolates nuclei away from membranous organelles and the cytosol
  2. Separates the soluble cytosolic compartment away from membranous organelles





# glycoRNAs gain access to the surface of living cells

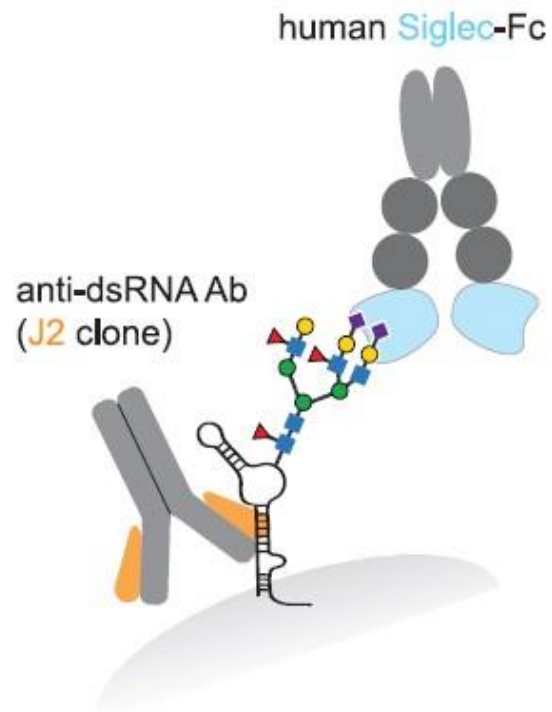
- ✓ done strictly at 40C to reduce or eliminate vesicular trafficking



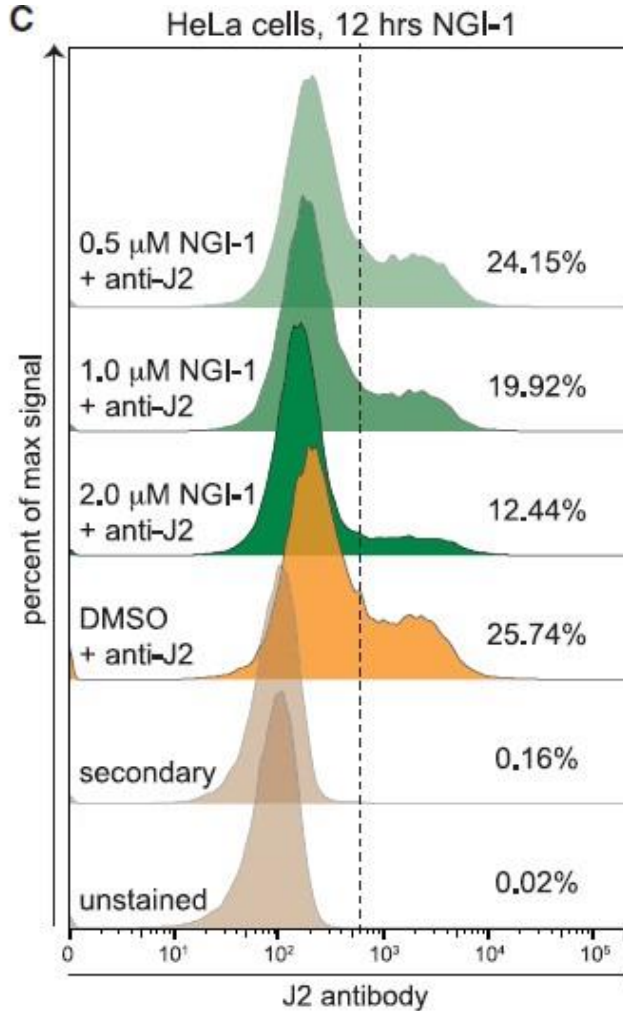
# Siglec receptors and anti-RNA antibodies recognize cell surface glycoRNAs

- ✓ Use antibodies that targeting RNA which have been associated with systemic lupus erythematosus (SLE)

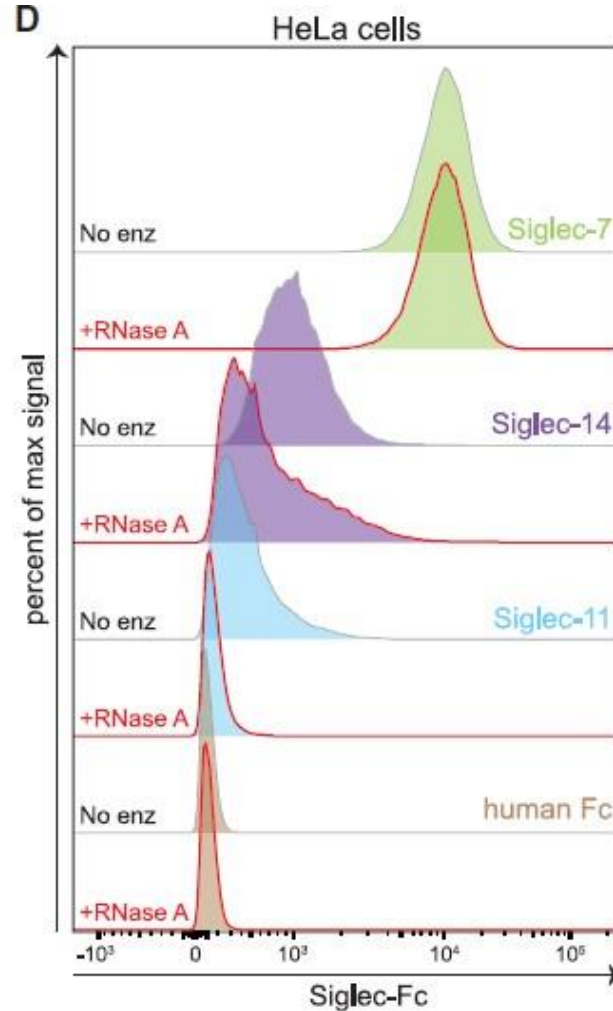
A



C



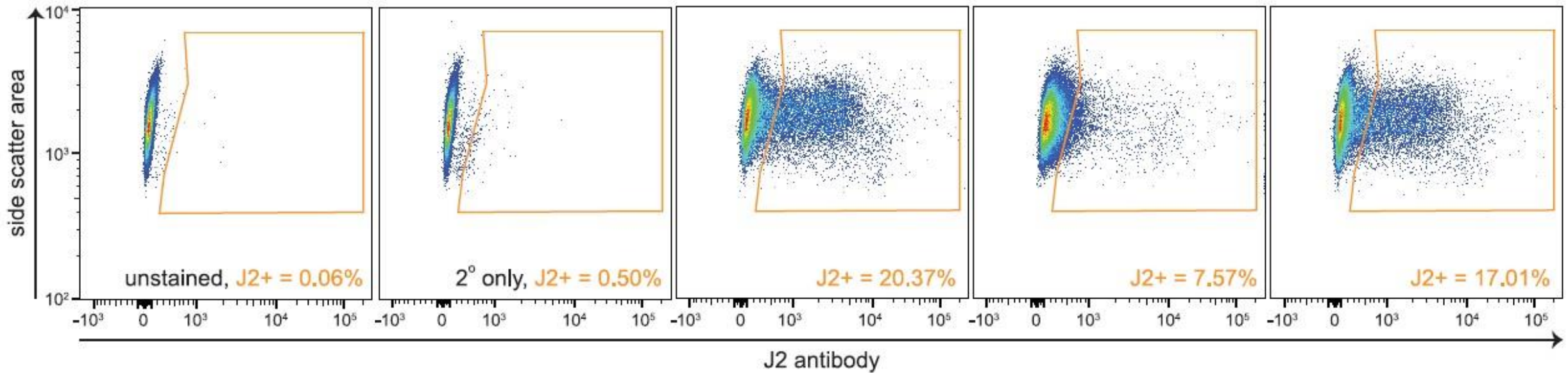
D



- ✓ Approximately 20% of a population of cultured HeLa cells showed positivity with J2 staining

**B** HeLa cells FACS:

J2 antibody	-	-	+	+	+
RNase A	-	-	-	+	+
RNase Inh.	+	+	+	-	+



## Chemical linkage of RNA to glycan

- ✓ Although the precise nature of the glycan-RNA linkage has not yet been determined, we speculate that direct glycosylation of native RNA bases is unlikely.
- ✓ The observed sensitivity to PNGase F, which cleaves the glycosidic linkage between asparagine and the proximal GlcNAc of N-glycans, implies an amide bond-containing linker that native nucleobases lack.
- ✓ It is possible that a precursor guanosine modification is necessary to establish an asparagine-like functionality capable of modification by OST (Oligosaccharyltransferase), or that a preassembled N-glycan carrier moiety is attached to nucleobases by some other chemistry.
- ✓ These possibilities are consistent with sedimentation of glycoRNAs in the sucrose gradient, which suggests a linker with a relatively small molecular weight.

# Limitations of study

- ✓ A major focus of the work presented leverages selective metabolic labeling of sialic acid with Ac4ManNAz.
- ✓ Because not all glycans contain sialic acid, it is possible that glycoform's beyond those reported here may also be conjugated to RNAs
- ✓ The precise linkage between the RNA template and carbohydrate remains unknown